## **Laboratory Testing for Pertussis**

Test	Sensitivity	Specificity	Comments
DFA	30-71%	66-100%	Should be used only as a screening
			test in conjunction with culture and
			PCR
Culture	11-73%	100%	Organism is extremely labile; requires
			prompt transport and plating;
			sensitivity best when specimen
			collected during catarrhal stage of the
			disease
PCR	61-99%	88-100%	No single technique is universally
			accepted or validated; no FDA cleared
			test available; culture should also be
			performed whenever PCR testing is
			performed
Serological Tests			Difficult to interpret, especially in
			immunized people; no FDA cleared
			test available; lack of standardized
			interpretive criteria
IgM			Generally not helpful
IgG	63%*	99%*	Specificity depends on the antigen
			used (PT is specific); more sensitive
			than IgA; paired sera can also be
			tested
IgA			Specificity depends on the antigen
			used; rarely detected in uninfected
			vaccinees; young infants may not be
			able to make IgA

## References:

- 1. Centers for Disease Control and Prevention. 2000. Guidelines for the Control of Pertussis Outbreaks. CDC Atlanta, GA.
- 2. Chan EL, et al. 2002. The Use of TaqMan PCR Assay for Detection of *Bordetella pertussis* Infection from Clinical Specimens. Arch Pathol Lab Med. 126: 173-176
- 3. Loeffelholz MJ, Thompson CJ, Lon, KS, Gilchrist MJR. 1999. Comparison of PCR, Culture, and Direct Fluorescent-Antibody Testing for Detection of *Bordetella pertussis*. J Clin Micro 37: 2872-2876
- 4. McGowan KL. 2002. Diagnostic Tests for Pertussis: Culture vs. DFA vs. PCR. Clin Micro Newsletter 24: 143-150
- 5. Muller F-MC, Hoppe JE, vonKonig C-HW. 1997. Minireview: Laboratory Diagnosis of Pertussis: State of the Art in 1997. J Clin Micro 35: 2435-2443

<sup>\*</sup>Massachusetts State Laboratory Institute's test, the only serological test recognized by CDC as confirmatory and only for patients in Massachusetts